THE FUNCTIONAL POTENTIAL OF CORAL-ASSOCIATED MICROBIAL **COMMUNITIES IN PSEUDOPTEROGORGIA AMERICANA FROM LA** PAGUERA, PUERTO RICO

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Coral-associated microbial communities are increasingly recognized as important components of the coral holobiont that influence coral health and disease. It has been suggested that these communities contribute to the health of the coral ecosystem through energy and nutrient cycling; others suggest that microbial communities provide corals with protection against infectious disease. To date however, there are few studies that directly address the functional role of these communities. In the present study, we examined the functional potential of the coral-associated microbial community found in the surface mucopolysaccharide layer (SML) of a common Caribbean coral, Pseudopterogorgia americana. The SML samples were collected in March of 2006 off the coast of La Paguera, Puerto Rico. DNA was extracted and amplified for use in a functional gene array, the GeoChip, which contains 24,000 probes targeting 10,000 functional genes involved in approximately 150 functional groups associated with biogeochemical processes. We examined the presence of these genes in community DNA from the SML of healthy P.americana colonies. Our preliminary data indicate that this microbial community possesses 1189 genes known to play a role in numerous biogeochemical processes. These processes include carbon degradation/fixation (159 genes), dissimilatory sulfite reduction (94 genes), metal homeostasis (203 genes), methane generation and oxidation (50 genes), nitrogen fixation/reduction (145 genes), ammonification (94 genes), and organic chemical degradation (444 genes). Our data, although preliminary, suggests that the coral-associated microbial community found in the SML of *P. americana* may play an active role in maintaining the coral holobiont through cycling of key nutrients, metals and organic contaminants. In on-going studies, we are evaluating the functional potential of healthy as well as diseased *P. americana* samples in triplicate using both SML and tissue samples.

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The mucous samples were collected in March of 2006 off the coast of La Paguera, Puerto Rico and stored in 5M GIT Buffer at -20C. The community DNA was extracted using a CTAB/Phenol/Chloroform protocol and amplified using the Amersham Biosciences TempilPhi amplification kit (25-6400-01).

Gene Category	# Genes	% Total	Example Genes
Carbon Degradation	126	10.60	cellulase, chitinase, laccase
Carbon Fixation	33	2.78	cdhE, cooF, ACSAB
Dissimilatory Sulfite Reduction	94	7.91	dsrA, dsrB
Metal Homeostasis	203	17.07	Ni, Cd, Co, Zn, Pb, Al, Hg
Methane Generation	19	1.60	mcr, mcrA, mcrG
Methane Oxidation	31	2.61	mmo, mmoA, pmo, pmoA
Nitrogen Fixation	48	4.04	nifD, nifH
Ammonification, Ammonia Assimilation	94	7.91	amoA, amoB, amoC, gdh
Nitrogen Nitrate/ Nitrite Reduction	97	8.16	nir, nirK, nirS, norB, narB
Organic Chemical Degradation	444	37.34	benzene, parathion
	Carbon Degradation Carbon Fixation Dissimilatory Sulfite Reduction Metal Homeostasis Methane Generation Methane Oxidation Nitrogen Fixation Ammonification, Ammonia Assimilation Nitrogen Nitrate/ Nitrite Reduction	Carbon Degradation126Carbon Fixation33Dissimilatory Sulfite Reduction94Metal Homeostasis203Methane Generation19Methane Oxidation31Nitrogen Fixation48Ammonification, Ammonia Assimilation94Nitrogen Nitrate/ Nitrite Reduction97	Carbon Degradation12610.60Carbon Fixation332.78Dissimilatory Sulfite Reduction947.91Metal Homeostasis20317.07Methane Generation191.60Methane Oxidation312.61Nitrogen Fixation484.04Ammonification, Ammonia Assimilation947.91Nitrogen Nitrate/ Nitrite Reduction978.16

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